

# Transcriptional regulation of bone sialoprotein gene by CO<sub>2</sub> laser irradiation

Yoko Sasaki<sup>1)</sup>, Shuang Wang<sup>1,2)</sup> and Yorimasa Ogata<sup>1,3)</sup>

<sup>1)</sup>Department of Periodontology, Nihon University School of Dentistry at Matsudo, Chiba, Japan

<sup>2)</sup>Stomatology College of Tianjin Medical University, Tianjin, China

<sup>3)</sup>Research Institute of Oral Science, Nihon University School of Dentistry at Matsudo, Chiba, Japan

(Received 2 December 2010 and accepted 24 January 2011)

**Abstract:** Bone sialoprotein (BSP), an early marker of osteoblast differentiation, has been implicated in the nucleation of hydroxyapatite during *de novo* bone formation. Low-power laser irradiation has a stimulating effect on cells and tissues. Although the carbon dioxide (CO<sub>2</sub>) laser is a hard surgical laser, we have attempted to use it at low energy density to achieve biological alterations. To investigate the effects of CO<sub>2</sub> laser irradiation on BSP gene transcription, we used rat osteoblast-like ROS17/2.8 cells. BSP mRNA levels were increased at 12 h after irradiation with the CO<sub>2</sub> laser (2 W, 20 s). Transient transfection assays using various sizes of the rat BSP gene promoter linked to the luciferase reporter gene showed that CO<sub>2</sub> laser irradiation induced luciferase activity of a -116 to +60 BSP promoter construct (pLUC3) at 12 h in the cells. Transcriptional stimulation by CO<sub>2</sub> laser irradiation was abrogated in the pLUC3 construct containing a 2-bp mutation in the fibroblast growth factor 2 response element (FRE). Gel shift analyses showed that CO<sub>2</sub> laser irradiation increased the binding of nuclear protein to FRE. These studies demonstrate that CO<sub>2</sub> laser irradiation increases BSP transcription via FRE in the rat BSP gene promoter. (J Oral Sci 53, 51-59, 2011)

**Keywords:** bone sialoprotein; CO<sub>2</sub> laser irradiation; osteoblast; transcription.

Correspondence to Dr. Yorimasa Ogata, Department of Periodontology, Nihon University School of Dentistry at Matsudo, 2-870-1 Sakaecho-nishi, Matsudo, Chiba 271-8587, Japan  
Tel: +81-47-360-9362  
Fax: +81-47-360-9362  
E-mail: ogata.yorimasa@nihon-u.ac.jp

## Introduction

Bone sialoprotein (BSP) is a mineralized tissue-specific non-collagenous glycosylated, phosphorylated and sulfated protein expressed almost exclusively in mineralized connective tissues (1-3). BSP is also expressed in breast, lung, thyroid and prostate cancers that metastasize to bone (4-7) and is associated with the formation of ectopic hydroxyapatite microcrystals in tumor tissues (8,9). Studies aimed at clarifying the transcriptional regulation of BSP have characterized the rat, mouse and human BSP gene promoters (10-12). These promoters have an inverted TATA box (-24 to -19) (13) and an inverted CCAAT box (-50 to -46), which are required for basal transcription (14). In addition, a cAMP response element (CRE; -75 to -68) (15), a fibroblast growth factor 2 response element (FRE; -92 to -85) (15,16), a pituitary-specific transcription factor-1 (Pit-1) motif (-111 to -105) (17), a homeodomain protein binding site (HOX; -199 to -192) (18,19), a transforming growth factor- $\beta$  (TGF- $\beta$ ) activation element (-499 to -485) (20), and a glucocorticoid response element (-920 to -906), overlapping an AP-1 site (-921 to -915) (1,21) have also been characterized.

There are several kinds of hard laser, such as the neodymium:yttrium-aluminum-garnet (Nd:YAG), erbium:YAG (Er:YAG) and carbon dioxide (CO<sub>2</sub>) lasers, and soft lasers such as the helium-neon and semiconductor lasers. These lasers are currently applied for a wide variety of dental procedures (22). Various effects, such as a bactericidal action in root canals and periodontal pockets, relief of pain, enhancement of wound healing, and stimulation of both fibroblast proliferation and macrophage phagocytosis have been reported (23,24). Low-power laser

irradiation exerts stimulatory effects on cell and tissues (25,26). After being absorbed, laser energy is transformed into heat, and therefore it would be expected that lasers would have useful effects such as anti-inflammation, pain elimination, wound healing and bone regeneration (27-31). Low-power lasers have also become well established in dentistry because of their anti-inflammatory, analgesic and regenerative effects, as well as a conditioning effect on tooth enamel (25,32). Early *in vivo* studies of bone regeneration after low-power laser treatment demonstrated increased bone deposition after tooth extraction, suggesting enhancing effects of this type of laser on ossification (33). These findings are supported by *in vitro* studies showing that low-power lasers significantly increased the proliferation, bone nodule formation and alkaline phosphatase activity of rat calvarial cells (34), and enhanced the osteogenic differentiation of murine mesenchymal stem cells (35). In the present study, we investigated the effects of a CO<sub>2</sub> laser at low energy density on BSP gene transcription in osteoblast-like cells.

## Materials and Methods

### Materials

Alpha minimum essential medium ( $\alpha$ -MEM), fetal calf serum (FCS), lipofectamine, penicillin, streptomycin, and TrypLE™ Express were obtained from Invitrogen (Carlsbad, CA, USA). The protein kinase C inhibitor H7 and protein kinase A inhibitor H89 were from Seikagaku Corporation (Tokyo, Japan). Tyrosine kinase inhibitor herbimycin A (HA) was purchased from Wako Pure Chemical Industries, Ltd. (Tokyo, Japan). PGL3-basic luciferase plasmid, pSV- $\beta$ -galactosidase control vector ( $\beta$ -Gal) and MAP kinase kinase (ERK1/2) inhibitor U0126 were purchased from Promega Co. (Madison, WI, USA). c-Jun N-terminal kinase (JNK) inhibitor SP600125 was from Calbiochem (San Diego, CA, USA). PP1 was from Biomol Research Laboratories, Inc. (Plymouth Meeting, PA, USA). A Quikchange Site-directed Mutagenesis Kit was obtained from Stratagene (La Jolla, CA, USA). All chemicals used were of analytical grade. A CO<sub>2</sub> laser transmitter (GC NANOLASER GL-III) was obtained from GC (Tokyo, Japan).

### Cell culture

Rat osteoblast-like ROS 17/2.8 cells were cultured at 37°C in 5% CO<sub>2</sub> and 95% air in  $\alpha$ -MEM containing 10% FCS. The cells were grown to confluence in 60-mm tissue culture dishes and then cultured in  $\alpha$ -MEM without serum for 12 h and then either left untreated or irradiated with the CO<sub>2</sub> laser at a dose of 0, 0.5, 1, 1.5, or 2 W for 20 s (power density: 0.5 w, 20 s; 0.357 J/cm<sup>2</sup>, 1 w, 20 s; 0.715

J/cm<sup>2</sup>, 1.5 w, 20 s; 1.07 J/cm<sup>2</sup>, 2 w, 20 s; 1.43 J/cm<sup>2</sup>), followed by incubation for 12 h, and followed the time course (3, 6, 12 and 24 h) after irradiation with the CO<sub>2</sub> laser (2 W, 20 s). Laser irradiation was performed at a distance of 300 mm (area of spot: 27.978 cm<sup>2</sup>) from the probe to the cell layer. Total RNA was extracted with guanidinium thiocyanate as described previously (18). RNA was isolated from triplicate cultures and analyzed for the expression of BSP and glyceraldehyde-3-phosphate dehydrogenase (GAPDH) mRNA by Northern hybridization, as described below.

### Northern hybridization

Twenty-five-microgram aliquots of total RNA were fractionated on a 1.2% agarose gel and transferred onto a Hybond N+ membrane. Hybridizations were performed at 42°C with <sup>32</sup>P-labeled rat BSP and GAPDH cDNA probes. Following hybridization, membranes were washed four times for 5 min each time at 21°C in 300 mM sodium chloride, 30 mM trisodium citrate, pH 7.0, containing 0.1% SDS. This was followed by two 20-min washes at 55°C in 15 mM sodium chloride, 1.5 mM trisodium citrate, pH 7.0, and 0.1% SDS. The hybridized bands, representing the rat BSP mRNA (two polyadenylated forms: 1.6 and 2.0 kilobases) and OPN mRNA were scanned in a Bioimaging analyzer (Fuji BAS 2500, Tokyo, Japan).

### Transient transfection assays

Exponentially growing ROS 17/2.8 cells were used for transient transfection assays. Twenty-four hours after plating, cells at 50-70% confluence were transfected using lipofectamine reagent. The transfection mixture included 1  $\mu$ g of a luciferase (LUC) construct (pLUC1, -18~+60; pLUC2, -43~+60; pLUC3, -116~+60; pLUC4, -425~+60; pLUC5, -801~+60) and 2  $\mu$ g of  $\beta$ -Gal vector as an internal control. Two days after transfection, the cells were deprived of FCS for 12 h and then irradiated with the CO<sub>2</sub> laser (2 W, 20 s), followed by culture for 12 h prior to harvest. The luciferase assay was performed in accordance with the supplier's protocol (picaGene, Toyo Inki, Tokyo, Japan) using a luminescence reader (Acuu FLEX Lumi 400; Aloka, Tokyo, Japan) to measure luciferase activity. The protein kinase inhibitors H7 (5  $\mu$ M) and H89 (5  $\mu$ M) were used to inhibit protein kinases C and A. HA (1  $\mu$ M) and PP1 (10  $\mu$ M) were used for tyrosine kinase and Src tyrosine kinase inhibition, respectively. U0126 (5  $\mu$ M) was used as an ERK1/2 inhibitor, and SP600125 (10  $\mu$ M) was used as a c-Jun N-terminal kinase (JNK) inhibitor. Oligonucleotide-directed mutagenesis by PCR was utilized to introduce dinucleotide substitutions using the Quikchange Site-directed Mutagenesis Kit. All constructs

were sequenced as described previously to verify the fidelity of the mutagenesis.

### Gel mobility shift assays

Confluent ROS 17/2.8 cells were irradiated with the CO<sub>2</sub> laser (2 W, 20 s) and incubated for 3 to 12 h in  $\alpha$ -MEM without FCS to prepare nuclear extracts, with the addition of extra proteinase inhibitors (the extraction buffer was 0.42 M NaCl, 1.5 mM MgCl<sub>2</sub>, 0.2 mM EDTA, 1 mM dithiothreitol, 25% (v/v) glycerol, 0.5 mM phenylmethylsulfonyl fluoride, 2  $\mu$ g/ml pepstatin A, and 1  $\mu$ g/ml aprotinin, pH 7.9). For gel shift analysis, double-stranded oligonucleotides encompassing the inverted CCAAT (nt -61 to -37, 5'-CCGTGACCGTGATTGGCTGCTGAGA), CRE (nt -84 to -59, 5'-CCCACAGCCTGACGTCGCACCGGCCG) and FRE (nt -98 to -79, 5'-TTTTCTGGTGAGAACCACA) in the rat BSP promoter were prepared by Bio-Synthesis, Inc. (Lewisville, TX). These double-stranded oligonucleotides were end-labeled with [ $\gamma$ -<sup>32</sup>P]ATP and T4 polynucleotide kinase. Nuclear protein extracts (3  $\mu$ g) were incubated for 20 min at room temperature with 0.1 pM radiolabeled double-stranded oligonucleotide in buffer containing 50 mM KCl, 0.5 mM EDTA, 10 mM Tris-HCl, pH 7.9, 1 mM dithiothreitol, 0.04% Nonidet P-40, 5% glycerol, and 1  $\mu$ g of poly (dI-dC). Following incubation, the protein-DNA complexes were resolved by electrophoresis on 5% non-denaturing acrylamide gels (38:2 acrylamide/bis acrylamide) run at 200 V at room temperature. After electrophoresis, the gels were dried, and autoradiograms were prepared and analyzed using an image analyzer.

### Statistical analysis

Quadruplicate samples were analyzed for each experiment, and experiments were replicated to ensure the consistency of the responses to drugs. Significant differences between the control and treatments were determined using unpaired Student's *t*-test.

## Results

### Effects of CO<sub>2</sub> laser irradiation on BSP mRNA expression

To study the effect of the CO<sub>2</sub> laser on BSP transcription, we performed Northern hybridization analysis of total RNA extracted from osteoblastic ROS 17/2.8 cells. First, the dose-response relationship of BSP induction by the CO<sub>2</sub> laser was established by treating the ROS 17/2.8 cells with different power outputs from the laser for 20 s and then incubating them for 12 h. CO<sub>2</sub> laser irradiation increased the levels of BSP mRNA at 0.5, 1, 1.5 and 2 W and had a maximal effect at 2 W (Fig. 1A). To determine

the time course of BSP mRNA expression, therefore, the cells were irradiated with the laser at 2 W for 20 s. It was found that laser irradiation up-regulated the level of BSP mRNA at 12 h after irradiation, whereas no effect on GAPDH was observed (Fig. 1B).

### CO<sub>2</sub> laser stimulation of rat BSP gene transcription

To determine how CO<sub>2</sub> laser irradiation affects BSP transcription, transient transfection analyses were performed using chimeric constructs encompassing different regions of the rat BSP gene promoter ligated to a luciferase reporter gene (pLUC1~5) transfected into ROS 17/2.8 cells. After CO<sub>2</sub> laser irradiation (2 W, 20 s), luciferase activities of BSP promoter constructs (pLUC3; nts -116~+60, pLUC4; nts -425~+60, pLUC5; nts -801~+60) were increased at 12 h (Fig. 2). Included within the DNA sequence, which

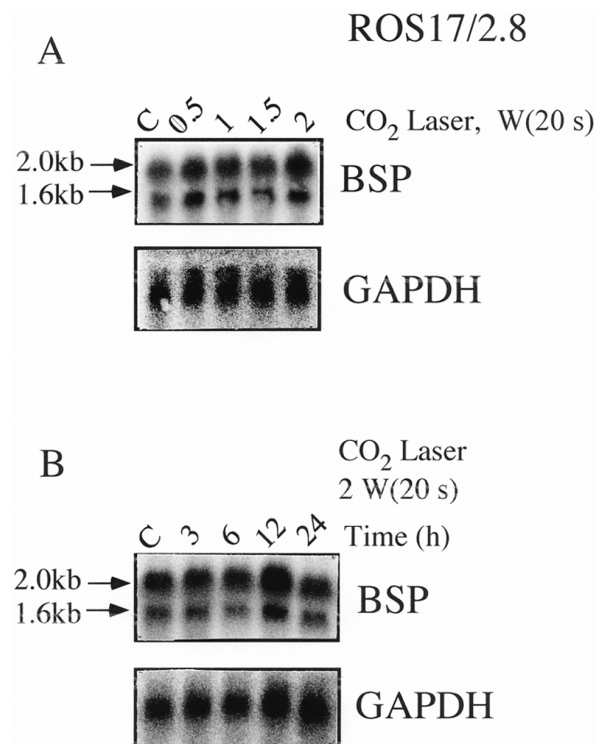


Fig. 1 Effect of CO<sub>2</sub> laser irradiation on BSP mRNA levels in ROS17/2.8 cells. A: Dose response effects of CO<sub>2</sub> laser irradiation (2 W, 20 s) on BSP mRNA levels. B: Time course effects of laser irradiation on BSP expression. Total RNA was extracted with guanidinium thiocyanate, and Northern hybridization was performed with <sup>32</sup>P-labeled rat BSP and GAPDH cDNA probes. The experiments were performed in triplicate for each data point. Results of representative hybridization analysis of control and CO<sub>2</sub> laser-treated cells are shown.

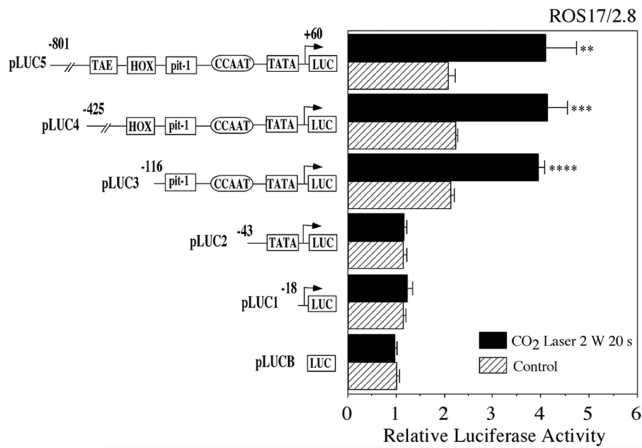


Fig. 2 Effect of CO<sub>2</sub> laser irradiation on BSP promoter activity in ROS 17/2.8 cells. Transient transfection of ROS 17/2.8 cells that were untreated or CO<sub>2</sub> laser (2 W)-irradiated for 20 s and cultured for 12 h was performed to determine the transcriptional activities of chimeric constructs that included various regions of the rat BSP promoter ligated to a luciferase reporter gene. The data for transcriptional activity obtained from four separate transfections with the constructs pLUC basic (pLUCB) and pLUC1 to pLUC5 have been combined, and the values are expressed with standard errors. \*\*( $P < 0.05$ ), \*\*\*( $P < 0.02$ ), \*\*\*\*( $P < 0.01$ ).

is unique in these regions, is an inverted CCAAT box (nts -50 to -46), a CRE motif (nts -75 to -68), a Runx2(1) motif (nts -84 to -79), a FRE motif (nts -92 to -85) and a Pit-1 motif (nts -111 to -105) (Fig. 3). To determine more precisely the target sites between nts -116 and -43 in the rat BSP gene promoter through which the CO<sub>2</sub> laser effect is mediated, we prepared a series of 5'-deletion constructs. The transcriptional activities of -84, -108 and -116BSPLUC in ROS17/2.8 cells were stimulated by the CO<sub>2</sub> laser (2 W, 20 s) at 12 h (Fig. 4). To determine the signaling pathways mediating the effects of the CO<sub>2</sub> laser, we tested the effects of several protein kinase inhibitors on the BSP transcription elicited by the laser. The tyrosine kinase inhibitor HA, the Src tyrosine kinase inhibitor PP1, and the ERK1/2 inhibitor U0126 abolished the effects of CO<sub>2</sub> laser irradiation on pLUC3 transcriptional activity (Fig. 5). To confirm the functional elements, we performed luciferase analyses using mutation constructs. After introducing 2-bp mutations into the CCAAT, CRE, FRE and Pit-1 elements within pLUC3, the basal activities of these constructs were lower than the basal transcriptional activity of pLUC3. The transcriptional stimulation elicited by the CO<sub>2</sub> laser (2 W, 20 s) was abrogated in the M-FRE construct (Fig. 6).

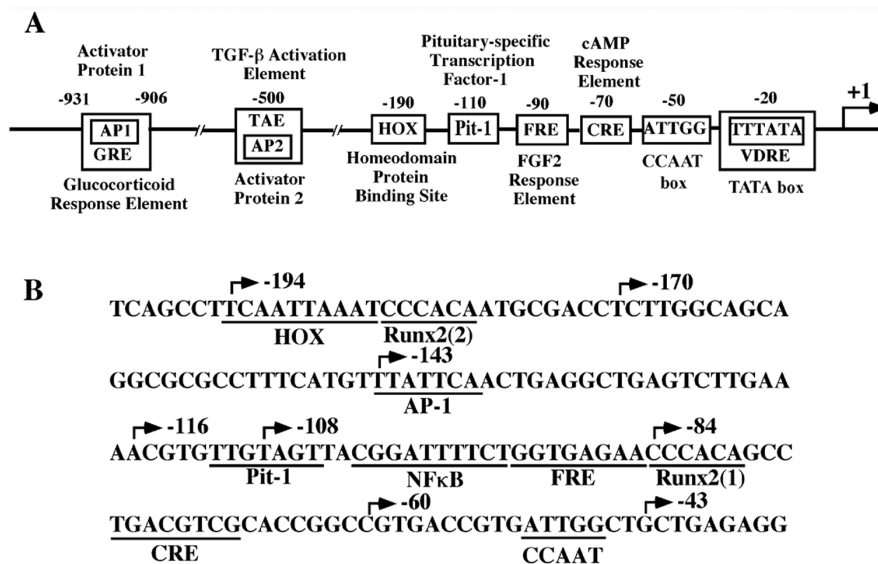


Fig. 3 Regulatory elements in the proximal promoter of the rat BSP gene. A: Diagram showing the positions of the inverted TATA and CCAAT boxes, vitamin D response element (VDRE) that overlaps the inverted TATA box, CRE (cAMP response element), FRE (FGF2 response element), Pit-1 (mediating the stimulatory effects of parathyroid hormone, PTH), the homeodomain protein-binding site (HOX), TAE (TGF- $\beta$  activation element) overlapping with AP2, and GRE (glucocorticoid response element) overlapping with AP1. B: The nucleotide sequence of the rat BSP gene proximal promoter, encompassing an inverted CCAAT box, and the CRE, Runx2(1), FRE, NF $\kappa$ B, AP1, Runx2(2) and HOX elements is shown from -201 to -35.

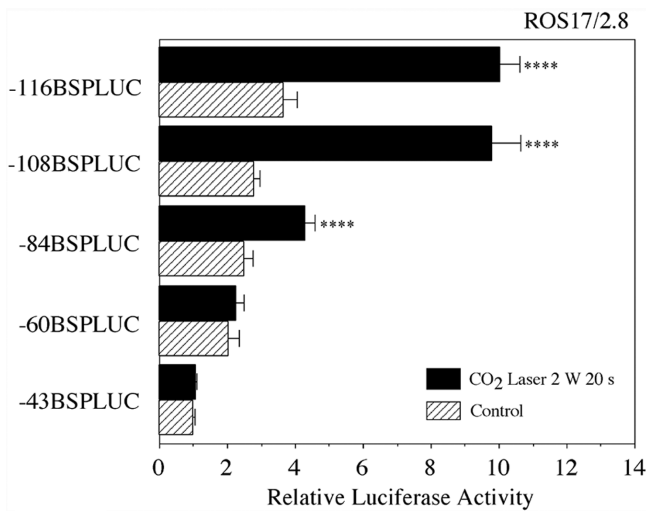


Fig. 4 Fine 5'-deletion mapping of the nts -116 to -43 elements in the BSP promoter. A series of rat BSP promoter 5'-deletion constructs were analyzed for relative promoter activity after transfection into ROS17/2.8 cells, which were subsequently irradiated with the CO<sub>2</sub> laser (2 W, 20 s) and incubated for 12 h. The results of transcriptional activity obtained from four separate transfections with the constructs -43 BSPLUC (-43 to +60), -60 BSPLUC (-60 to +60), -84 BSPLUC (-84 to +60), -108 BSPLUC (-108 to +60), and -116 BSPLUC (-116 to +60) have been combined, and the values expressed with standard errors. \*\*\*\*( $P < 0.01$ ).

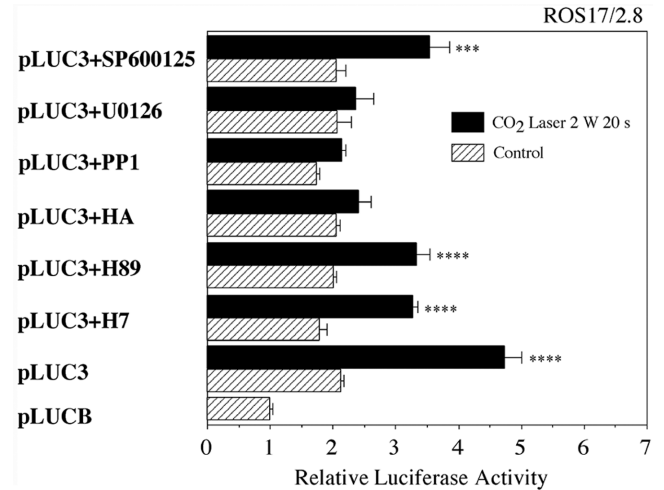


Fig. 5 Effects of kinase inhibitors on transcriptional activation by CO<sub>2</sub> laser irradiation. Transient transfection analysis of pLUC3 in ROS 17/2.8 cells irradiated with the CO<sub>2</sub> laser (2 W, 20 s) and incubated for 12 h is shown together with the effects of a PKC inhibitor (H7, 5  $\mu$ M), PKA inhibitor (H89, 5  $\mu$ M), tyrosine kinase inhibitor (herbimycin A; HA, 1  $\mu$ M), MEK inhibitor (U0126, 5  $\mu$ M), Src tyrosine kinase inhibitor (PP1, 10  $\mu$ M), and JNK inhibitor SP600125 (10  $\mu$ M). The results of transcriptional activity obtained from four separate transfections with the constructs were combined and the values expressed with standard errors. Significant differences from control: \*\*\*( $P < 0.02$ ), \*\*\*\*( $P < 0.01$ ).

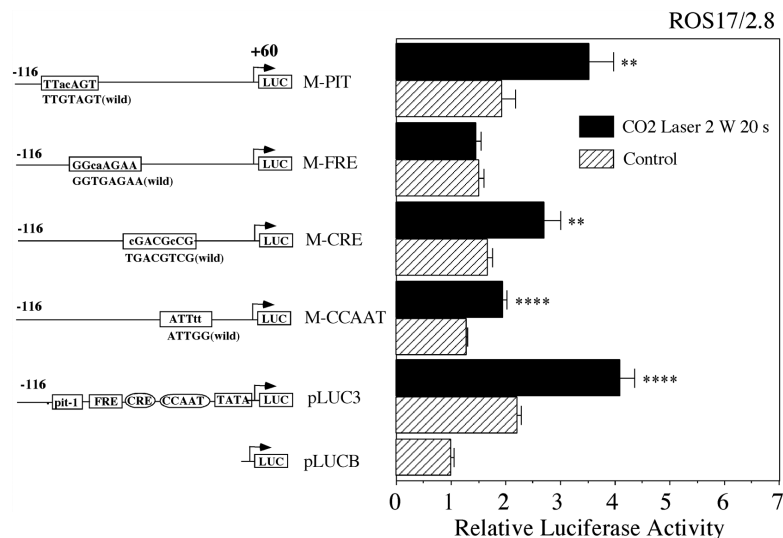


Fig. 6 Site mutation analysis of luciferase activities. Dinucleotide substitutions were made within the context of the pLUC3 promoter fragment. M-CCAAT (ATTt), M-CRE (cGACGcCG), M-FRE (GGcaAGAA) and M-PIT (TTAcAGT) constructs were analyzed for relative promoter activity after transfection into ROS17/2.8 cells, and examined for regulatory effects after irradiation with the CO<sub>2</sub> laser (2 W, 20 s) and incubation for 12 h. The data for transcriptional activity obtained from four separate transfections with the constructs were combined, and the values expressed with standard errors. \*\*( $P < 0.05$ ), \*\*\*\*( $P < 0.01$ ).

## Gel mobility shift assays

To identify nuclear proteins that bind to the CCAAT, CRE and FRE elements and mediate the effects of the CO<sub>2</sub> laser on transcription, double-stranded oligonucleotides were end-labeled and incubated with equal amounts (3 μg) of nuclear proteins extracted from confluent ROS 17/2.8 cells that were either untreated (control) or treated with the CO<sub>2</sub> laser (2 W, 20 s) for 3, 6 and 12 h. When we used the inverted CCAAT elements as probes, the DNA-protein complexes were not changed by laser irradiation (Fig. 7, lanes 1-4). When FRE was used as a probe, irradiation with the laser increased the binding of nuclear proteins to FRE at 3 h, and this was followed by a return to the basal level at 6 h and a further increase at 12 h (Fig. 7, lanes 9-12). Irradiation with the laser did not induce formation of the CRE-protein complex (Fig. 7, lanes 5-8). To further characterize the proteins in the complex formed with the FRE, we used antibodies against several transcription

factors. Anti-Dlx5 and anti-Msx2 antibodies disrupted FRE-binding proteins (Fig. 8, lanes 4 and 5), while anti-Runx2 and anti-Smad1 antibodies did not reduce FRE-protein complex formation (Fig. 8, lanes 6 and 7).

## Discussion

In this study, we showed that CO<sub>2</sub> laser irradiation enhanced transcription of the rat BSP gene by targeting FRE in its proximal promoter. BSP is a valuable marker of osteogenic differentiation and bone formation, and the RGD cell attachment sequence facilitates attachment of normal bone or cancer cells to mineralized tissue surfaces. BSP is also highly expressed in osteotropic cancer cells and plays a role in tumor metastasis to bone (3,8,9). BSP and OPN, as members of the family of small integrin binding proteins (SIBLINGs), are the major constituents

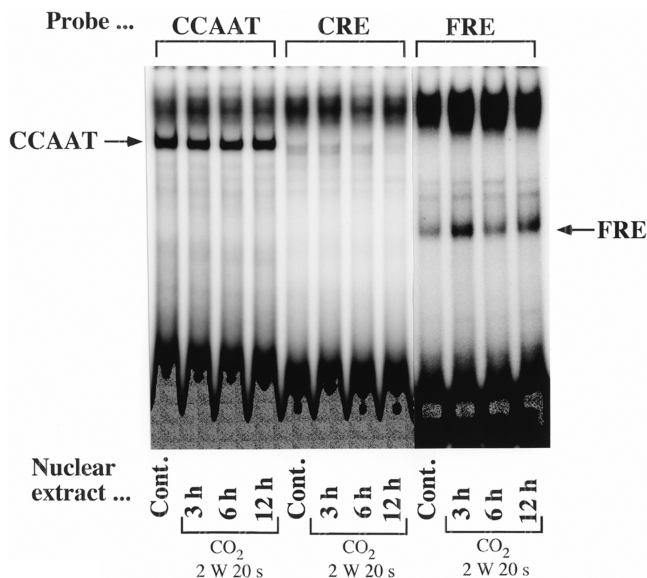


Fig. 7 Increase in the binding of nuclear proteins to the FRE induced by CO<sub>2</sub> laser irradiation. Radiolabeled double-stranded inverted CCAAT (-61 CCGTGACCGTGATT GGCTGCTGAGA -79), CRE (-84 CCCACAAGCCT GACGTTCGACCGGCCG -59) and FRE (-98 TT TTCTGGTGAGAACCCACA -79) oligonucleotides were incubated with nuclear protein extracts (3 μg) obtained from ROS17/2.8 cells that were untreated or irradiated (lanes 1, 5 and 9) with the CO<sub>2</sub> laser (2 W, 20 s) and incubated for 3 h (lanes 2, 6 and 10), 6 h (lanes 3, 7 and 11), or 12 h (lanes 4, 8 and 12). DNA-protein complexes were separated on 5% polyacrylamide gel in low-ionic-strength Tris-borate buffer, dried under vacuum, and exposed to an imaging plate for quantitation using an image analyzer.

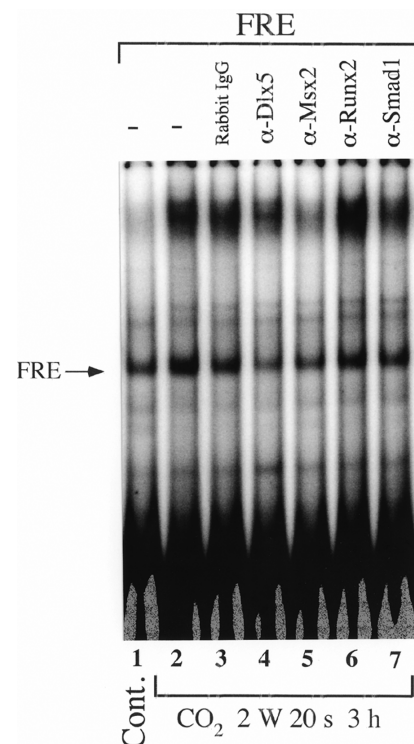


Fig. 8 Specific binding of nuclear proteins to the FRE. Radiolabeled double-stranded FRE was incubated with nuclear protein extracts (3 μg) obtained from ROS17/2.8 cells that were untreated (lane 1) or irradiated with the CO<sub>2</sub> laser (2 W, 20 s) and cultured for 3 h (lanes 2-7). Supershift experiments were performed with 0.4 μg of antibodies against IgG (lane 3), Dlx5 (lane 4), Msx2 (lane 5), Runx2 (lane 6) and Smad1 (lane 7) added separately to each gel shift reaction.

of non-collagenous proteins in bone matrix, and expression of their gene products has been shown to be under the control of calcitropic hormones and cytokines (3,8). In the present study, CO<sub>2</sub> laser irradiation increased the level of BSP mRNA and transcriptional activity of the BSP gene (Fig. 1 and 2), thus supporting the possibility that CO<sub>2</sub> laser irradiation could be utilized to regulate osteoblastic function in the local bone environment.

Using transient transfection assays, we localized the CO<sub>2</sub> laser-responsive region to the proximal promoter (pLUC3, -116 to -43) of the rat BSP gene, which encompasses an inverted CCAAT box, and the CRE, Runx2(1), FRE and Pit-1 motifs (Fig. 3). However, no further increase in transcription of the pLUC4 and pLUC5 constructs was observed (Fig. 2), suggesting that CO<sub>2</sub> laser irradiation influences BSP transcription through the response element in pLUC3 that contains the FRE. Using fine 5'-deletion constructs between nts -116 to -43 in the rat BSP promoter, -84BSPLUC, containing CRE and Runx2(1), was identified as one of the targets of CO<sub>2</sub> laser irradiation. Luciferase activity of -108BSPLUC was further enhanced by laser irradiation, suggesting that the responsive region lies not only in -84BSPLUC but also in the -108BSPLUC construct (Figs. 3 and 4). Transcriptional activity was totally abrogated by mutation of FRE in the pLUC3 construct (Fig. 6), thus further confirming that FRE is the functional element responsible for the effect of CO<sub>2</sub> laser irradiation on BSP transcription.

The involvement of FRE was further supported by gel shift analyses (Fig. 7). Transcription factors that bind specifically to FRE were demonstrated by supershift assays, which indicated that anti-Dlx5 and anti-Msx2 antibodies interacted with FRE-binding proteins (Fig. 8).

Animal studies have demonstrated that low-power laser irradiation enhances the functional attachment of titanium implants to bone and promotes bone healing and mineralization (36). As the regeneration of bone defects is a key problem in regenerative medicine, the biostimulatory effects of low-power lasers could be an attractive form of co-treatment for repair of such defects (37). The CO<sub>2</sub> laser is easily absorbed by water, and therefore irradiated deep tissues are rarely injured (38). Although the CO<sub>2</sub> laser is a hard surgical laser, we have attempted to use it at low energy density to achieve biological alterations (31). Laser irradiation has been reported to stimulate the proliferation of osteoblast-like cells (39), and a similar effect of low-output CO<sub>2</sub> laser irradiation on fibroblasts has also been reported (40).

In this study, we have shown that the CO<sub>2</sub> laser can enhance BSP gene transcription through the tyrosine kinase, Src tyrosine kinase and ERK1/2 signaling pathways,

and identified FRE in the rat BSP gene promoter as the target of CO<sub>2</sub> laser-mediated enhancement of BSP gene transcription.

## Acknowledgments

This work was supported in part by a Grant-in-Aid for Scientific Research (C; No. 22592319), a grant from the Supporting Project for Strategic Research in Private Universities from the Ministry of Education, Culture, Sports, Science, and Technology, Japan (MEXT), 2008-2012, a grant from the Strategic Research Base Development Program for Private Universities from the Ministry of Education, Culture, Sports, Science, and Technology, Japan (MEXT), 2010-2014 (S1001024), and a Nihon University Multidisciplinary Research Grant for 2009 and 2010.

## References

1. Ogata Y, Yamauchi M, Kim RH, Li JJ, Freedman LP, Sodek J (1995) Glucocorticoid regulation of bone sialoprotein (BSP) gene expression: identification of a glucocorticoid response element in the bone sialoprotein gene promoter. *Eur J Biochem* 230, 183-192.
2. Tye CE, Rattray KR, Warner KJ, Gorden JAR, Sodek J, Hunter GK, Goldberg HA (2003) Delineation of the hydroxyapatite-nucleating domains of bone sialoprotein. *J Biol Chem* 278, 7949-7955.
3. Ogata Y (2008) Bone sialoprotein and its transcriptional regulatory mechanism. *J Perio Res* 43, 127-135.
4. Bellahcène A, Merville MP, Castronovo V (1994) Expression of bone sialoprotein, a bone matrix protein, in human breast cancer. *Cancer Res* 54, 2823-2826.
5. Bellahcène A, Maloujahmoum N, Fisher LW, Pastorino H, Taqliabue E, Ménard S, Castronovo V (1997) Expression of bone sialoprotein in lung cancer. *Calcif Tissue Int* 61, 183-188.
6. Bellahcène A, Albert V, Pollina L, Basolo F, Fisher LW, Castronovo V (1998) Ectopic expression of bone sialoprotein in human thyroid cancer. *Thyroid* 8, 637-641.
7. Waltregny D, Bellahcène A, Riet IV, Fisher LW, Young M, Fernandez P, Déwe W, Leval JD, Castronovo V (1998) Prognostic value of bone sialoprotein expression in clinically localized human prostate cancer. *J Natl Cancer Inst* 90, 1000-1008.
8. Ganss B, Kim RH, Sodek J (1999) Bone sialoprotein. *Crit Rev Oral Biol Med* 10, 79-98.

9. Waltregny D, Bellahcène A, Leval XD, Florkin B, Weidle U, Castronovo V (2000) Increased expression of bone sialoprotein in bone metastases compared with visceral metastases in human breast and prostate cancers. *J Bone Miner Res* 15, 834-843.
10. Li JJ, Sodek J (1993) Cloning and characterization of the rat bone sialoprotein gene promoter. *Biochem J* 289, 625-629.
11. Benson MD, Aubin JE, Xiao G, Thomas PE, Franceschi RT (1999) Cloning of a 2.5kb murine bone sialoprotein promoter fragment and functional analysis of putative *Osf2* binding sites. *J Bone Miner Res* 14, 396-405.
12. Kim RH, Shapiro HS, Li JJ, Wrana JL, Sodek J (1994) Characterization of the human bone sialoprotein (BSP) gene and its promoter sequence. *Matrix Biol* 14, 31-40.
13. Li JJ, Kim RH, Sodek J (1995) An inverted TATA box directs downstream transcription of the bone sialoprotein gene. *Biochem J* 310, 33-40.
14. Kim RH, Sodek J (1999) Transcription of the bone sialoprotein gene is stimulated by v-Src acting through an inverted CCAAT box. *Cancer Res* 59, 565-571.
15. Samoto H, Shimizu E, Matsuda-Honjyo Y, Saito R, Nakao S, Yamazaki M, Furuyama S, Sugiya H, Sodek J, Ogata Y (2003) Prostaglandin E2 stimulates bone sialoprotein (BSP) expression through cAMP and FGF2 response elements in the proximal promoter of the rat BSP gene. *J Biol Chem* 278, 28659-28667.
16. Shimizu-Sasaki E, Yamazaki M, Furuyama S, Sugiya H, Sodek J, Ogata Y (2001) Identification of a novel response element in the rat bone sialoprotein (BSP) gene promoter that mediates constitutive and fibroblast growth factor 2-induced expression of BSP. *J Biol Chem* 276, 5459-5466.
17. Ogata Y, Nakao S, Kim RH, Li JJ, Furuyama S, Sugiya H (2000) Parathyroid hormone regulation of bone sialoprotein (BSP) gene transcription is mediated through a pituitary specific transcription factor-1 (Pit-1) motif in the rat BSP gene promoter. *Matrix Biol* 19, 395-407.
18. Nakayama Y, Nakajima Y, Kato N, Takai, H, Kim DS, Arai M, Mezawa M, Araki S, Sodek J, Ogata Y (2006) Insulin-like growth factor-I increases bone sialoprotein (BSP) expression through fibroblast growth factor-2 response element and homeodomain protein-binding site in the proximal promoter of the BSP gene. *J Cell Physiol* 208, 326-335.
19. Benson MD, Bargeon JL, Xiao G, Thomas PE, Kim A, Cui Y, Franceschi RT (2000) Identification of a homeodomain binding element in the bone sialoprotein gene promoter that is required for its osteoblast-selective expression. *J Biol Chem* 275, 13907-13917.
20. Ogata Y, Niisato N, Furuyama S, Cheifetz S, Kim RH, Sugiya H (1997) Transforming growth factor- $\beta$ 1 regulation of bone sialoprotein gene transcription: identification of a TGF- $\beta$  activation element in the rat BSP gene promoter. *J Cell Biochem* 65, 501-512.
21. Yamauchi M, Ogata Y, Kim RH, Li JJ, Freedman LP, Sodek J (1996) AP-1 regulation of the rat bone sialoprotein gene transcription is mediated through a TPA response element within a glucocorticoid response unit in the gene promoter. *Matrix Biol* 15, 119-130.
22. Ishikawa I, Aoki A (2006) Laser in periodontics. In: Carranza's clinical periodontology, 10th ed, Newman MG, Takei HH, Klokkevold PR, Carranza FA eds, Saunders, St Louis 1035-1038.
23. Walsh LJ (1997) The current status of low level laser therapy I dentistry. Part 1. Soft tissue applications. *Aust Dent J* 42, 247-254.
24. Bedenarska K, Rozga B, Kolodziejczyk K, Szosland D, Leyko W, Bryszewska M (1998) Effect of low-power red light laser irradiation on the viability of human skin fibroblast. *Radiat Environ Biophys* 37, 215-217.
25. Kon K (1992) In-vivo study of low-power laser irradiation on new bone formation of rat carvaria defect. *Nippon Laser Shigakkai Shi* 3, 35-44. (in Japanese)
26. Gomi K, Yashima A, Shirakawa S, Kanazashi M, Sekine A, Kobatashi K, Nagano T, Matui K, Yamaguchi H, Arai T (2005) Effects on mineralization by low-level irradiation of hard laser in three-dimension cultured rat bone marrow cells. *Nippon Laser Shigakkai Shi* 16, 10-15. (in Japanese)
27. Kasai S, Yamamoto Y, Kotani H, Ando N, Kamiya K, Kono T (1997) The basis of the low power laser irradiation for wound healing. *Nippon Laser Igakkai Shi* 18, 27-34. (in Japanese)
28. Suzuki H, Tanaka H, Hashimoto K (1994) The study on the effect of diode laser on cultured osteoblastic cells. *Nippon Laser Shigakkai Shi* 5, 41-46. (in Japanese)
29. Nakashima M, Hirata K, Shinoda K, Watanabe K, Terano T, Hoji T, Sekie I, Kimura K, Mukouyama Y (1991) Analgesic effect of soft-laser irradiation. Part1. Effect of soft-laser irradiation on rat jaw opening reflex. *Nihon Shika Hozongaku Zasshi* 34,



- 329-334. (in Japanese)
30. De Long HP, Schakenraad JM, Busscher HJ (1988) Spreading of human fibroblast on pretreated human root dentin. *J Biol Buccale* 16, 31-34.
  31. Nagasawa A, Asai H, Yamamoto H, Kata K (1987) Experimental study on bone healing activation effect. *Nippon Laser Igakkai Shi* 7, 73-74. (in Japanese)
  32. Turhani D, Scheriau M, Kapral D, Benesch T, Jonke E, Bantleon HP (2006) Pain relief by single low-level laser irradiation in orthodontic patients undergoing fixed appliance therapy. *Am J Orthod Dentofacial Orthop* 30, 371-377.
  33. Takeda Y (1988) Irradiation effect of low-energy laser on alveolar bone after tooth extraction. Experimental study in rats. *Int J Oral Maxillofac Surg* 17, 388-391.
  34. Ozawa Y, Shimizu N, Kariya G, Abiko Y (1998) Low-energy laser irradiation stimulates bone nodule formation at early stages of cell culture in rat calvarial cells. *Bone* 22, 347-354.
  35. Abramovitch-Gottlib L, Gross T, Naveh D, Geresh S, Rosenwaks S, Bar I, Vago R (2005) Low level laser irradiation stimulates osteogenic phenotype of mesenchymal stem cells seeded on a three-dimensional biomatrix. *Lasers Med Sci* 20, 138-146.
  36. Khadra M, Rønold HJ, Lyngstadaas SP, Ellingsen JE, Haanaes HR (2004) Low-level laser therapy stimulates bone-implant interaction: an experimental study in rabbits. *Clin Oral Implants Res* 15, 325-332.
  37. Stein E, Koehn J, Sutter W, Wendtlandt G, Wanschitz F, Thurnher D, Baghestanian M, Turhani D (2008) Initial effects of low-level laser therapy on growth and differentiation of human osteoblast-like cells. *Wien Klin Wochenschr* 120, 112-117.
  38. Riggs K, Keller M, Humphreys TR (2007) Ablative laser resurfacing: high-energy pulsed carbon dioxide and erbium:yttrium-aluminum-garnet. *Clin Dermatol* 25, 462-473.
  39. Ueda Y, Shimizu N (2003) Effects of pulse frequency of low-level laser therapy (LLLT) on bone nodule formation in rat calvarial cells. *J Clin Laser Med Surg* 21, 271-277.
  40. Katada H (1994) The effect of laser irradiation on secretion of PGE2 from gingival fibroblasts. *Aichi-Gakuin Daigaku Shigakkai Shi* 32, 159-167. (in Japanese)